

Product Sheet

H_TNFRSF9(4-1BB) Reporter 293 Cell line

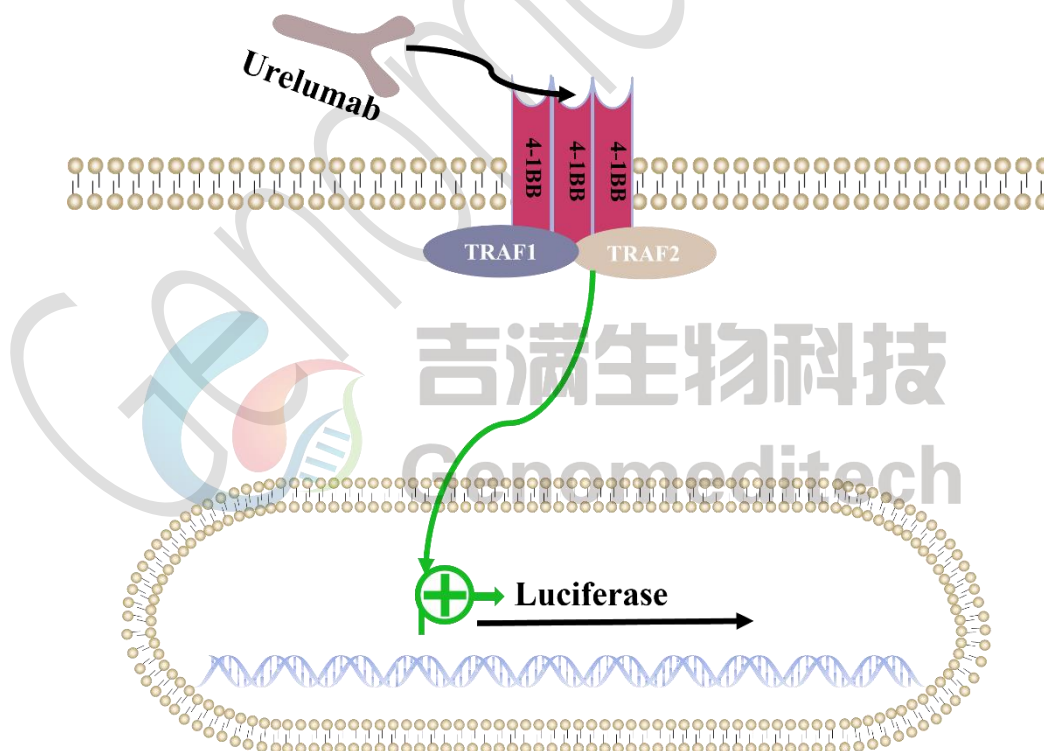
Catalog number: GM-C04832

Version 3.3.1.250320

4-1BB (CD137) is a protein in the TNF receptor superfamily, mainly found on T cells and NK cells. It is crucial for immune responses, enhancing T cell proliferation, survival, and function. Its activation is mediated by the ligand 4-1BBL (CD137L), expressed on activated dendritic cells, B cells, and some tumor cells. The signaling pathway involves TRAF and NF- κ B, promoting cell survival and proliferation.

Activation of 4-1BB recruits TRAF2 and TRAF1, activating downstream pathways like NF- κ B and MAPK, leading to cytokine production (e.g., IL-2 and IFN- γ). This enhances T cell immune responses, supporting anti-tumor immunity and infection clearance. Thus, 4-1BB is a key target in cancer immunotherapy, with potential as an immune checkpoint inhibitor.

H_TNFRSF9(4-1BB) Reporter 293 Cell line is a clonal stable cell line constructed using lentiviral technology, constitutive expression of the TNFRSF9(4-1BB) gene, along with signal-dependent expression of a luciferase reporter gene. When 4-1BBL binds to 4-1BB, it activates downstream signaling pathways, leading to the expression of luciferase. The luciferase activity measurement indicates the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to TNFRSF9(4-1BB).



Specifications

Quantity	5E6 Cells per vial, 1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Recovery Medium	DMEM+10% FBS+1% P.S
Growth medium	DMEM+10% FBS+1% P.S+4 µg/mL Blasticidin+0.75 µg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO ₂

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ GM-040404
Puromycin	Genomeditech/ GM-040401
Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio)	Genomeditech/ GM-87876MAB
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/ GM-040503

Figures

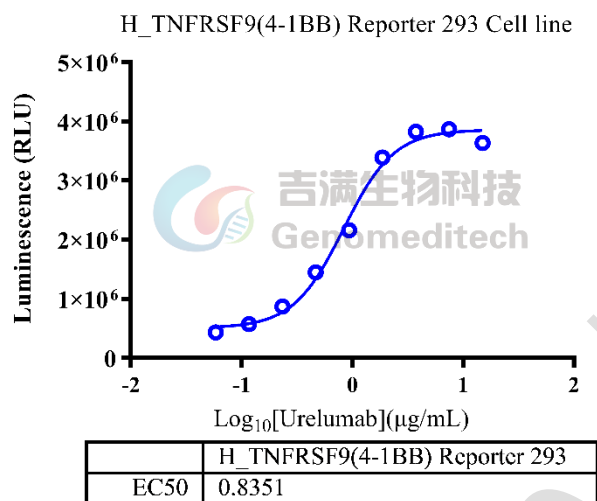


Figure 1 | Response to Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio). The H_TNFRSF9(4-1BB) Reporter 293 Cell line (Cat. GM-C04832) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio) (Cat. [GM-87876MAB](#)) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The maximum induction fold was approximately [8.7]. Data are shown by drug mass concentration.

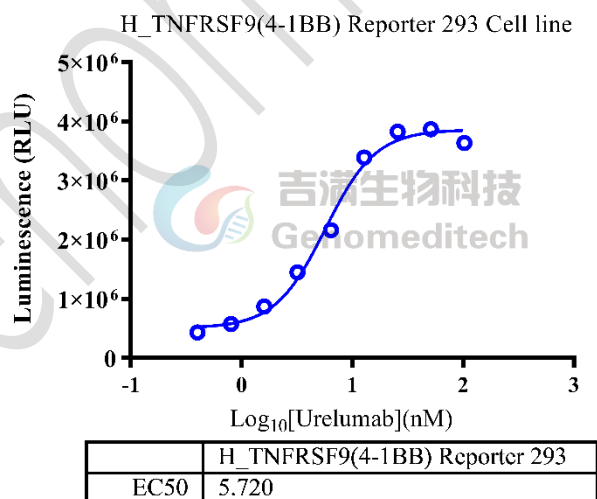


Figure 2 | Response to Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio). The H_TNFRSF9(4-1BB) Reporter 293 Cell line (Cat. GM-C04832) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio) (Cat. [GM-87876MAB](#)) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The maximum induction fold was approximately [8.7]. Data are shown by molar mass concentration.

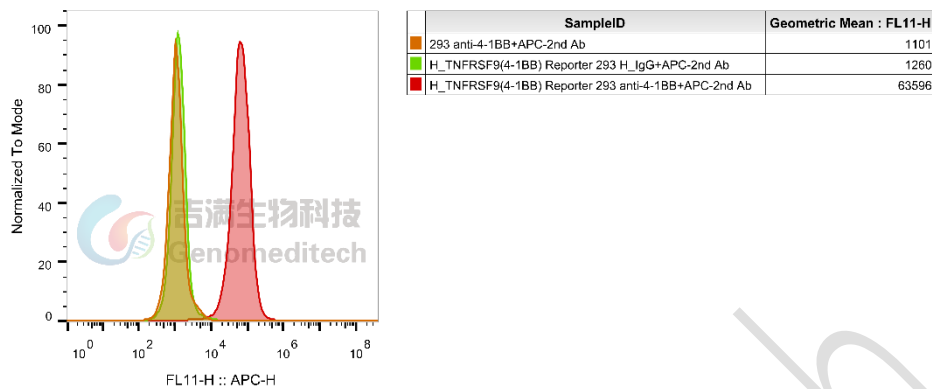


Figure 3 | H_TNFRSF9(4-1BB) Reporter 293 Cell line (Cat. GM-C04832) was determined by flow cytometry using Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio) (Cat. [GM-87876MAB](#)).

Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C . Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately $176 \times g$ for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at $176 \times g$ for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5×10^6 cells/mL.
- Aliquot 1 mL into each vial.
- Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM+10% FBS+1% P.S+4 µg/mL Blasticidin+0.75 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.
- b) Remove and discard culture medium.
- c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).
- e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended

Medium Renewal: Every 2 to 3 days

Notes

- a) Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.
- b) Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

Related Products

4-1BB	
H_TNFRSF9(4-1BB) Reporter Jurkat Cell line	Cynomolgus_TNFRSF9(4-1BB) CHO-K1 Cell Line
H_TNFRSF9(4-1BB) CHO-K1 Cell Line	
Anti-H_4-1BB hIgG2 Antibody(Utomilumab)	Anti-H_TNFRSF9(4-1BB) hIgG4 Antibody(Urelumab)
Anti-TNFRSF9(4-1BB) hIgG4 Reference Antibody (Urelbio)	
CD3	
Jurkat CD3-BsAb Reporter Cell Line	Cynomolgus_CD3 HEK-293 Cell Line
Cynomolgus_CD3E(Membrane Bound ECD) CHO-K1 Cell Line	H_CD3 CHO-K1 Cell Line
H_CD3 HEK-293 Cell Line	H_CD3E(Membrane Bound ECD) CHO-K1 Cell Line
Mouse_CD3 HEK-293 Cell Line	
Anti-CD3 epsilon hIgG1 Antibody [OKT-3 (muromonab)]	Anti-CD3 hIgG1 Antibody(CH2527)

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